## Digestion Method for Soils

#### Purpose

Digest oven-dry soil to produce solution with total elements in organic form for analysis.

#### Preparation:

Sulfuric/salicylic acid solution:

- 1. Weigh out 34 g of salicylic acid on plastic weigh boat.
- 2. Gently pour into 1 L flask.
- 3. Add 18 M (concentrated) sulfuric acid to flask (but not totally to 1 L)
- 4. Add magnetic stir bar and gently stir and heat (on 2) until fully dissolved.
- 5. Top with stopper and cover in tin foil (to keep it in the dark) to cool overnight or until room temperature.
- 6. When cool, remove stir bar.
- 7. Fill to 1 L with sulfuric acid and store in amber bottle in refrigerator.

Weigh oven-dry soil

- 1. Place acid washed 100 mL digestion tubes in metal rack.
- 2. Use autoclave pencil to write numbers 1-40 on the tubes. You will use this to identify your samples later.
- 3. Weigh  $0.5000 \pm 0.0025$  g of dried and ground soil and record weights in data sheet along with tube number.
- 4. Include 2 blanks, 4 reps, and 2 NAPT soil standard (or equivalent).
- 5. Add three Hengar boiling chips to each tube.
- 6. If leaving overnight, cover the top with aluminum foil to avoid particles from floating into the tubes.

### Digestion:

- 1. Turn on the digestion block about 30 minutes before the digestion is to begin.
- 2. Place the rack with weighed plant tissue in the fume hood and on a metal baking sheet.
- 3. Pipet in 5 mL of  $H_2SO_4/Salicylic$  acid to each tube.
- 4. Pipet 2 mL of  $H_2O_2$  0.5 mL at a time (Fisher H325-500) in 0.5 mL increments.
  - a. Add 0.5 mL, vortex (on high), add 0.5 mL, vortex, add 0.5 mL, vortex, add 0.5 mL, vortex
- 5. Put the tubes on the block when it is under 300 °C (250 °C for the first phase, 290 °C for remaining phases).
- 6. Set scrubber to high, turn on scrubber. After 10 min, set scrubber to medium and keep it here until step 10.
- 7. Place the manifold on top of the tubes.
- 8. The tubes heat on the block at 320 °C for 15 minutes.
- 9. Remove rack from the block (keep manifold and scrubber on) and place back on the baking sheet.
- 10. Cool for 20 minutes.

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- 11. Gently remove the manifold and place on a second baking sheet in the hood.
- 12. Add 1.5 mL  $H_2O_2$  to each tube and vortex.
- 13. Repeat steps 7-11.
- 14. Add 1.5 mL  $H_2O_2$  to each tube and vortex.
- 15. Repeat steps 7-11.
- 16. Add 1.0 mL of  $H_2O_2$ , vortex.
- 17. Repeat steps 7-11.
- 18. Add 0.5 mL of  $H_2O_2$ , vortex.
- 19. Repeat steps 7-11.

# If the samples are not clear at this point, repeat steps 18 and 19, by adding 0.5 mL $H_2O_2$ and placing back on the block.

- 20. Turn off block completely
- 21. Once the tubes have cooled for at least 30 min, remove the manifold and set scrubber to low to suck the remaining acid from the manifold and tubing.
- 22. Slowly add ~75 mL of DI to each tube.
- 23. Allow the tubes to cool completely in the hood (overnight or for several hours). Cover with aluminum foil or place stoppers in the tubes to avoid anything from floating in.
- 24. Top off tubes to 100 mL calibration mark with DI water. Cover with a stopper and invert 5 times.
- 25. Allow the tubes to cool for at least an hour. Do not invert again!! The digestate should be clear, and there will be white gunk on the bottom (boiling chips).
- 26. Pour enough digestate to half fill a scintillation vial, close the vial with a cap and shake several times. You are cleaning the scintillation vial with the digestion solution. Pour this into the nalgene waste bucket.
- 27. Pour enough digestate into a scintillation vial to almost fill it, swirl and pour out.
  \*\*Make sure you empty the contents of the nalgene waste bucket into a labeled waste carboy when done.
- 28. **Collect your sample:** Fill a scintillation vial with the digestate (third quarter of the sample) and cap.
- 29. Samples are shelf-stable (do not need to be refrigerated or frozen). Keep in a cool, dry place until ready to analyze.

#### Waste Tag:

- water = 94%
- sulfuric acid = 4%
- Hengar boiling chips = 1%
- salicylic acid = 1%